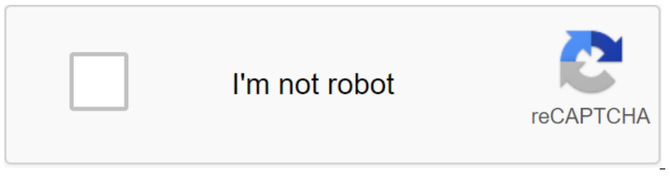

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Familia cannaceae pdf

Canna Canna? generalisTaxonom'aReino: PlantaeDivision: MagnoliophytaClase: MonocotyledoneaeSubclase: CommelinidaeOrden: TsichiberalesFamilia: Cannaceae Juss. n. 1) Gender: CannaL.Species See synonymy Following names are considered synonymous with Canna: Cannacorus Mill., Gard. Dict. Apr. 4: s.p. (1754). Catubala Adans., Pham. Pl. 2: 67 (1763). Xyphostylis Raph., F. Tellur. 4: 52 (1838). Distemon C.D. Boucher, Linnaeus 18: 494 (1845). Heurticus C.D. Boucher, Linnea 18: 485 (1845). Achirida Horan., Prodr. Monogra. Scitam.: 18 (1862). Wikidata edit Canna redirects here. For other purposes, see Canna (disambigation). Cannaceae (Cannaceae) are a family of herbaceous, perennial and rhizomes plants belonging to the order of zingiberales monocotilendons. The family has only one genus, Cannes, with twenty-three species known as achiras, adeiras or kanyas de India. They are tropical and subtropical plants native to America that are characterized by their distic leaves, their flowers with petal-shaped stamen with single tic and their staminodes that look like petals. Different types of Canna are grown for human nutrition in several regions of the world: in Mexico and the Antilles, in Res (from Colombia to Argentina), as well as in Hawaii, Australia, Vietnam, China and Indonesia. Canna stands out as the most famous, variable and distributed among these species. Some of them are grown for decorative purposes, especially in France, Spain, England, Hungary, other places in Europe and Brazil. The etymology and common name of the name of Canna comes from the Hebrew cane, which means cane. Common Spanish names for different types of Canna are very diverse, such as canna de las Indias, Platanillo, Platanillo de Cuba, Kane India, Kan choir, achira, achera, sago and dragon language. However, achira prevails in South America and the Crea of Indians in Spain. Description of the habit of growth and inflorescence in Canna indica var. Varshevich. Fruits in the formation of the achira with its characteristic prickly structures that cover it. Notice the persistent bowls on the fruit. These are perennial herbs and glabruses. They have two types of stem: one airy, without branching and straight; another underground, risomatosis and simpodial or monopod branch according to the species is considered to be with mucilage channels. Leaves alternate, distic (spiral), simple, whole stock, petiolate or more or less petiole-free, with a well-developed leaf, the middle vein of the sheet has airway channels, with pinnate veins (peni-parallel), shell on the base, without ligula polished or stypulous. They are differentiated into basal shells, petiol and foil. Pods overlap around the stem and support it. Flowers of hermaphrodite, without the plane of symmetry, epigia (with a hellish ovaries), often last one day. Perigonium consists of 3 separated and imbrikized chalicistic chalices (sometimes called external tepals) and 3 petals (sometimes called internal tapals) combined, as well as absorbed. Androceo consists of one fertile stump (back environment), a petal, with its thread attached to the staminoids and adnan to the investigation. Dust is non-toxic or also called monootek (i.e. it is the middle anther, the other half expands, forming a structure similar to petal and sterile), a two-group, subapic lateral position, longitudinal dehisection. Staminodes, usually 3 or 4, can be one and rarely 5, long, petals similar to fertile stamenas, 1 longer than the other, and repeated, all in their basal part connates and adnates to corolla. Pollen has no holes and its exina is very small. The dynasty consists of 3 wrists (the middle carpal is the anterix), connected together, with 3 locoles. The ovary withdrawal, outwardly papillous, with numerous eggs on the loculus, anatropes and bitegmic, the thought of the placenta. The style is terminal and laminar, flattened and more or less petal. Stigmatization is unique, extends along one side of the style. Nectarios are found in ovarian septos. Inflorescences are terminal and can be defined or uncertain. The central axis is triangular in cross section, with tristic bracts, each bract usually associated with small peaks, only 1 or 2 flowers. Secondary inflorescence represents if the straightened tyros is made up of thorns or a cluster of peaks. The fruit is usually a warts capsule that opens irregularly to the collapse of the fruit wall. The bowl is persistent in the fruit. The seeds are spherical, black, associated with the hair group (which is a modified ale). They have endosperm and perisperm and are rich in starch. The embryo is straight. The main chromosomal number is x-9. Canna Indica, Flowers and Seeds, Reunion Island, 2005. The t juventus variety of Ciiberales is represented in the global flora until the generic is edited by Kubicki (1998). In each region there are local flora, in the Spanish-speaking regions expected in Spanish, which, if found in the region, describe the zingiberales and the species of Cannacea is present in the region that flora covers, which can be consulted in institutions dedicated to botany with libraries available to the public, such as botanical gardens. Flora can be ancient and the latest species described in the region cannot be found in them, so consultations of the latest primary taxonomic literature (recent taxonomic monographs, taxonomic reviews and recent inventory (control data) of species and genera in the region) or with a local specialist who knows about them may be necessary. The latest volumes of the most modern flora usually follow a classification based on APG - they may have some differences - but many families as can be found in old flora and volumes have undergone significant changes in the groups that make up them or even in their taxonomic concept, so a comparison with a constituency like this may be necessary to synchronize them. The variety of Cannaceae will then be briefly considered. Its function is to have an image of each family while reading sections of ecology, phylogna and evolution. I don't go to The Environment of Cannacies is pro-natural in the tropical and subtropical regions of America, spreading from sea level to mountain slopes below 3,000 meters above sea level. One species (Canna indica) has been naturalized in tropical and subtropical regions of Asia and Africa. Many derivative interspecies species and hybrids are grown in gardens around the world. These are plants of moist bodies of water, river banks or wetlands, often growing in places up to 10 cm deep in the water. Pollen is deposited in style before the flower opens, directly in the stigma or slightly below it, so that many species are self-debuded (they are said to be autogamous). In non-amogam species, pollination biology is poorly studied, but the most common pollinators may be nectar bees, butterflies, moths and birds (hummingbirds). Seeds are often dissipated by water and remain viable for a long time. In fact, the seeds of about 600 years, found in a tomb on a South American archaeological site, sprouted and the plant flourished. Phylogenia and taxonomy filogenia Cannaceae monophilia is supported as molecular DNA analysis, so morphological analyses.ingiberales is as follows: Ciniberales Musaceae Strelitziaceae Heliconiaceae Core of Tsingiberales Tsinghiberasa Costaceaee Marantaceae Cannaceae Cannaceae Cannaceae Cann'ceas very closely related to marant'ceas, as seen from the treasure trove, with which he shares several unusual reproductive characteristics, such as floral asymmetry, reducing the number of fertile stamens to one dusty tree and secondary representation of pollen. Pollen grains are large and spherical and, like many other members of the scurvy, have a very small exina and a very extended layer of intine. Many hybrids of achira have mottled leaves, depicted a leaf of canna variegatifolia. The Taxonomy Family has been recognized by all plant classification systems, from the Engler system to Cronquist, Tahtayan, to the APG III system. Until a few years ago, most taxonomists recognized more than 50 species in the genus Cannes, but thanks to the work of Paulus Maas, based especially on the concepts of biogeographical history, the degree of interspecific hybridization during domestication and plasticity of morphological features, this number was reduced to just over twenty species. Such works were then completed by a taxonomic analysis of Nobuyuki Tanaka. The following list is based on the review of the Cannasei family's 2001 review of the Species of Asia and the New World, as well as for M. M. Ciciarelli for Argentine species in 1989, 25 with the addition of some post-publishing work 27 or the discovery of new species 27 or the discovery of new species, as in the case of Cannes dates back in 2007. Kanna Amabilis T. Koyama and Nob.Tanaka from Chaco Province, Argentina. Canna rises ciciar., from the province of Buenos Aires (Argentina). Cannes Bangui Kraenzl from Peru and Bolivia. Canna Coccinia Mill., from northern Argentina. Rosco compact canna, southern Brazil and northern Argentina. Canna bleach var. bleached (Lindl.) Nob.Tanaka, from Mexico to Colombia. Canna frees Goran from Brazil. Flaccid canned Salisb., from the southeastern United States, Central America and the Caribbean. Cannes Glauc L., distributed from South Carolina (United States) to tropical America. Canna points to L., 37, distributed in Mexico and Central America, cultivated and naturalized in various parts of the world. It is believed to have several taxonomic varieties: Canna indica var. Flava (Roscoe's former Baker) Nob. Tanaka. Canna points to var. waste paper (Hook.) Nob. Tanaka. Canna points to var. Santa Rosae (Krantzl.) Nob.Tanaka. Canna points to var. Varshevich (A.Dietr.) Nob.Tanaka. Canna iridiflora Ruiz and Pav., distributed in Costa Rica in Peru. Canna Jacobiniflora T. Koyama and Nob. Tanaka, from Currents in Argentina. Canna jaegeriana Urban, Caribbean and north of South America. Canna Liliflora Varsch. former Planch,, Bolivia. Canna panicata Ruiz and Pav., from tropical America. - Cimarrona achira from Peru , Canna Patens (Ayton) Roscoe, from southern Brazil. Canna Peduculata Sims, southern Brazil. Multitube canna T. Koyama and Nob. Tanaka, Jujui Province in Argentina. Canna speciosa Roscoe former Sims, from Brazil and northeastern Argentina. Canna Stenant Nob. Tanaka, distributed from Paraguay to the province of Miciones in Argentina. Canna Tuenkheim Krenzl. Canna variegatifolia Ciciar., from the province of Santa Fe, Argentina. Canna Generalis, photograph taken on the island of San Nicolas, Cape Verde, August 2007. Canna Liliflora. In addition to these species found in nature, all artificial interspecies hybrids to improve the color and size of the flower, as described in the next section, have received binomial names. Like this. In the early 20th century, Liberty Hyde Bailey detailed two types of garden achiras: C. x generalis and C. x orchiodes, to classify the decorative achirs cultivated at the time. Over time, the two groups intersected, so their differentiation became increasingly uncertain, and the two names given by Bailey became redundant (both were called decorative atyrams). For this reason, all current varieties are grouped in Notospecting Canna Generalis, and it classifies various varieties into cult groups, as confirmed by the International Code of Nomenclature of Cultivated Plants. The economic importance of achira rhizomes (especially those from Canna indica) is used as food, as their rhizomes are consumed fried or cooked as potatoes and from them excellent flour used in the production of cookies is obtained. Its seeds are used in the creation of necklaces and rosaries and its leaves serve to wrap certain products. The root decoction also serves as a diuretic substance, being the starch of the rhizome is very digestible and therefore beneficial for infants and sick people. In addition, in some places they use poultices made with roots prepared for emollient purposes. Finally, stems and leaves matter as fodder for cattle. However, the greatest importance of species and hybrids (e.g. Canna generalis) is their use as ornamental plants. Decorative use of the Origin of decorative achiras Interspecific hybridization played a fundamental and dominant role in decorative achirs. This was made possible by the large ecological differentiation of parental species, which determined that they lack barriers to hybridization and very good fertility as a result of hybrids. Differences in flower color between species of Canna are controlled by a large number of genes and their emulators, inhibitors, deadly. From recombination in interspecies hybrids that are segregated to so many genes, a large number of heterocytic genotypes have emerged with new colors and color combinations that can also continue to generate new genetic variability. This hybridization was also responsible for transgressive segregation, especially in the length and width of the staminod and the flier sizes that affected not only the height of the plant, but also the size of the flower. The effective vegetative spread of species and hybrids allowed, on the other hand, the fixation of the most beneficial genotypes, even if they had a high degree of infertility. Atirs created by Theodore Anne (hybrids between C. indica and C. glauca) and other enthusiasts in Germany and France (hybrids between C. iridiflora and C. warscwiczii) originated between 1848 and 1863. Although both types of hybrids were much more attractive than the parent species, the flowers were still very small. A great development was achieved in 1868 when the achiras called Crozy were released (their creator, French horticulturist Antoine Crosi), gladiolus or French dwarfs (collectively called C. generalis Bailey). This group originated from the interbreeding and reverse crossing between the first two groups and contained diploids, triploids and complex heterozygotes for chromosomal reordering. Later in this group, C. flacida, a new species was introduced, resulting in the created by 1872 the so-called Italian achiers, irises, orchids or giant flowers (grouped under the name C. orchiodes Bailey, which are now considered a simple synonym of C. ' generalis). Genetically these varieties were sterile desinaptic diploids or allotriploids. By the end of the 19th century, Carl Sprenger in Italy and Wilhelm Pfitzer in Germany continued to produce new products through crossbreeding and the introduction of new species. After interspecific hybridization, polyploidy is another important mechanism in the origin of chicory with more durable and durable colors. It is clear that the 44 years between 1848 and 1892 were a fruitful period in the genetic improvement of the achira, and where the choice is largely regulated by the following increased tolerance to cold, reduced height, inflorescences much higher than foliage, erect flowers, increase the size of the flower and a variety of colors and foliage. The trend in improving the achira in recent years has been the achievement of lower varieties with more air stems on the rhizomes. This type of plant is usually thick and very florist. It is noteworthy that the choice for the two main uses of achira concerned not only different organs (roots or flowers), but also different environments. While the selection of decorative achir took place in the mild climate of Europe, completely new to the genus, the selection for the production of starch in the rhizomes took place in their native habitat. It is interesting to note that two different screening targets in different habitats, both ended with triploids: the increase in floral sizes in ornamental achiras and the size of the rhizomes in achiras used to produce starch (Canna edulis, today considered synonymous with C. indica). The food use of sago, also called achira, arawak, imm, chisgua, maraca and capacho, is a species used as food (Canna edulis, a name that is considered synonymous with C. indica). Until the late 1950s, the sago was a constant diet in the diet of Andean communities. Sancochado was absorbed as a garrison and called this san training. Its main use was for the production of sago flour derived from the roots of chicory or sago plant, which was used as the main food for children in the first years of life in preparations with fresh cow's milk (sago atoll). Sago starch is high in amilaser, which is an important protein. It shows very high viscosity in the temperatures that are exposed in the production of pasta, making it easier to handle hot gels compared to other starches. It is a great source of nutrients for children, the elderly and people suffering from digestive problems. Baking requires 80% of production, the household uses 15%, industry 1%, and the rest for other purposes. In Colombia, for example, 800 hectares are planted with an average yield of 1,000 kilograms of flour per hectare, which is processed in hundreds of craft bakeries to produce the famous achira cake, which is a regional symbol in some regions of South America. Biscuits, muffins and sweets are also made in a hygienic package and excellent preparation and presentation. Unlike andean countries, where cultivation is declining, in a few years, the area of crops has increased to 30,000 hectares, and flour is used to make clear noodles (gluten noodles), popular food throughout Southeast Asia, at a lower price and the production of pasta of exceptional quality. In the food industry, chicory starch is consumed as cookies, meat starches, muffins as a thickener in instant soups and castings for children, in the canned food industry, in the production of sauces, as toppings in dietary products and in the production of sweet gums among others. In the pharmaceutical industry, it is widely used as fillers in the production of medicines in tablets. In the textile industry, starch clothing and to achieve adhesion fibers that make up clothing. In the paper and adhesive industry, chicory starch is unsympathetic and is not an obstacle to paper recycling. Medicinal use of several species of this genus have been used in folk medicine because of the various pharmacological actions presented by the various organs of these plants. Thus, canna risoma coccinea is used as diuretic, anti-asthmatic and emollient while the leaves are used as anti-reeumatic. The roots of Cannes glauc are used as diuretic and diaphoretic. Inflorescence of the chicory variety, New zealand. Achiers grow best when placed in full sun on well-drained, rich or sandy soils. Vegetative growth begins when the winter cold ends and progresses throughout the warm season until the first frosts occur, after which the air part of the plants dries up, and the underground rhizomes remains a reserve organ that will sleep (i.e. its kidneys remain dormant, without cell divisions and with very little metabolic activity) until the temperature begins to rise again. In mild winter areas, rhizomes can remain on the ground in winter. However, in regions with very wet winters, where, for example, the surface horizon of the soil is frozen to -10oC, the rhizomes should be removed from the ground as soon as autumn is finished and preserved, protected from low temperatures (more than 7oC), until spring. The roots should be planted 10 to 15 cm deep, separating them 50 cm from each other. During the growth of stems and leaves, plants can be fertilized with mineral nutrients (containing nitrogen, phosphorus, potassium and calcium) if the soil is not well supplied by them. Similarly, adequate humidity should be maintained throughout the growth period, as the achiers do not resist drought. Irrigation can be blurred or removed when the leaves begin to dry at the beginning of the cold season. Atirs can also be grown in pots or containers where precautions need to be taken when it comes to irrigation and consuming nutrients such as fertilizer. The fruit of the achira multiplication begins to open. Inside there are bright black seeds. Sexual reproduction: Although there are some species or populations that are automatic, most atyrs are illogical, meaning they must cross paths with different plants in order to give to descendants. The transport of pollen from one plant to another (pollination) is carried out by bats or hummingbirds, which are rewarded with nectar, which canna flowers produce in abundance. The resulting seeds, very difficult, should be stored before planting in hot water for 24 hours to soften the shell. Optimal planting season at the end of winter, on peat and sandy substrate and with temperatures of at least 18 to 20oC, which may be slightly smaller once the seeds have sprouted. The multiplication of seeds, due to sexual reproduction, is a method of increasing the populations of some natural species. Alogamia, combined with the gra variability of Canna varieties, prevents the characteristics of varieties from remaining unchanged when sexually multiplied due to genetic recombination. Therefore, to maintain plants with the same characteristics and spread to sterile varieties (many of them triploid), it is necessary to use vegetative reproduction. Asexual or vegetative reproduction: To multiply the asexual asexuality, rhizome division is used. They are cut into several pieces, so that each carries one or two vigorous buds. This operation can be done using the extraction and storage of rhizomes in winter in harsh climatic conditions or at the beginning of the warm season in a benign climate. The pieces are stored and stored moist, planting them when they begin to sprout, which happens when the temperature starts to rise. Once planted they should be fertilized and watered regularly. Micropropaganda methods are also used to spread certain varieties quickly. However, the achiras are quite recalcitrant, which will be grown in vitro. Diseases that can attack different species of Canna in the garden are few and can be grouped, according to their etiology, into diseases caused by bacteria, fungi or viruses. Diseases caused by bacteria: Highlights primarily kidney rot and shoots caused by bacteria Can't. Symptoms in the leaves are a series of spots that when merged give an aspect of scratches or list them. Symptoms of pimpollos are blackening and wet rot in inflorescences before flowering. The best control for this disease is to plant healthy rhizomes that do not come from plants with these symptoms. Fungal diseases: rot of stem and rhizomes caused by fungi of the genus Risoctonia and Fuzarium is important, which can colonize the roots, and then the base of the stems, causing withering and subsequent death of infected plants. Control of this disease is based on disinfecting the soil before planting and avoiding excess moisture or parting. Alternariasis is another important disease in achirah. It is caused by the fungus of the genus Alternaria. Achira rust is another disease caused by the fungus Puccinia cannacearum, which causes orange pustules on the leaves. Diseases caused by viruses: Achira plants can be attacked by different types of viruses, which determines the appearance of spotted, listed, dwarf and deformed plants or with a particularly distorted growth of leaves and flowers. The Chico mottle virus (CYMV from Canna yellow mottle badnavirus), which only affects species of the genus Canna. Bean is a yellow mosaic virus (BYMV) that infects achira, gladiolos, frisia and various legumes. The virus is a yellow striatum (CaYSV, canna yellow band virus), which causes yellow stretch marks and mosaics on the leaves. Pests Achira plants can be attacked by aphids that weaken the plant and transmit viruses, piglets (such as Aspidiotus lataniae), caterpillars of some butterflies (such as Gortyna ochracea or Xanthoecia flavago) that drill stalks and wire worms or white worms (Agriotes lineatus and anoxiaillo vsa) that devour the roots. Notes 86 in LAPG III 2009, references to Elspeth Haston, James E. Richardson, Peter F. Stevens, Mark W. Chase, David Harris. Linear Angiosperm Phylogeny Group (LAPG) III: Linear Family Sequence in APG III Botanical Journal of the Linnaeus Society, Volume 161, No. 2. (2009), page 128-131. doi:10.1111/j.1095-8339.2009.01000.x Key: citeulike:6006207 pdf: . Royal Horticultural Society, Kew. A detailed checklist for the Cannaceae family and the Canna family. b c Malaret, August (1970). Lexicon of fauna and flora. Madrid: Permanent Commission of the Association of Spanish Language Academies. 569 euros. Judd, W. S.; C. S. Campbell, E. A. Kellogg,. F. Stevens, M.J. 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